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#### **EUROPEAN PATENT APPLICATION**

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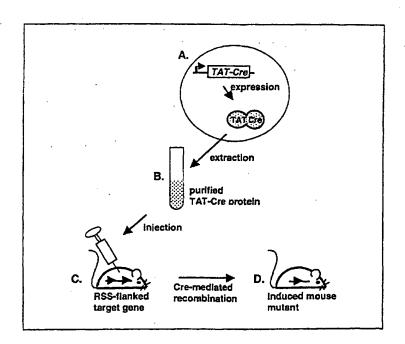
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- (54) Transduction of recombinases for inducible gene targeting
- (57) The present invention provides the use of a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain for pre-

paring an agent for inducing target gene alteration in a living organism, suitable fusion proteins and a method for the production of said fusion proteins.

Fig. 1



#### Descripti n

[0001] The present invention provides the use of a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain for preparing an agent for inducing target gene alteration in a living organism, suitable fusion proteins and a method for the production of said fusion proteins.

#### Background

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[0002] For some years targeted mutagenesis in totipotent mouse embryonic stem (ES) cells has been used to inactivate genes, for which cloned sequences were available (Capecchi Trends in Genetics 5, 70 - 76 (1989)). Since ES cells can pass mutations induced in vitro to transgenic offspring in vivo, it is possible to analyze the consequences of gene disruption in the context of the entire organism. Thus, numerous mouse strains with functionally inactivated genes ("knock out mice") have been created by this technology and utilized to study the biological function of a variety of genes. [0003] A refined method of targeted mutagenesis, referred to as conditional mutagenesis, employs a site-specific recombination system (e.g. Cre/loxP or Flp/frt - Sauer and Henderson, N. Proc. Natl. Acad. Sci. USA 85, 5166-5170 (1988); Senecoff et al., J. Mol. Biol., 201, 405 - 421 (1988)) which enables a temporally and/or spatially restricted alteration of target genes (Rajewsky et al, J. Clin. Invest., 98, 600 - 603 (1996)). The creation of conditional mouse mutants requires the generation of two mouse strains, i.e. the recombinase recognition strain and the recombinase expressing strain. The recombinase recognition strain is generated by homologous recombination in ES cells as described above except that the targeted exon(s) is (are) flanked by two recombinase recognition sequences (hereinafter "RRS"; e.g. loxP or frt). The type of recombination event mediated by the recombinase depends on the disposition of the RRS, with deletions, inversions, translocations and integrations being possible (Torres and Kühn, Oxford Univerity Press, Oxford, New York (1997)). By placing the RRS into introns, an interference with gene expression before recombination can be avoided. The recombinase expressing strain contains a recombinase transgene (e.g. Cre, Flp) whose expression is either restricted to certain cells and tissues or is inducible by external agents. Crossing of the recombinase recognition strain with the recombinase expressing strain recombines the RRS-flanked exons from the doubly transgenic offspring in a prespecified temporally and/or spatially restricted manner. Thus, the method allows the temporal analysis of gene function in particular cells and tissues of otherwise widely expressed genes. Moreover, it enables the analysis of gene function in the adult organism by circumventing embryonic lethality which is frequently the consequence of gene mutation. For pharmaceutical research, aiming to validate the utility of genes and their products as targets for drug development, inducible mutations provide an excellent genetic tool. However, the current systems for inducible recombinase expression in transgenic animals suffer from a certain degree of leakyness in the absence of the inducer (Kühn et al., Science 269(5229):1427-9 (1995); Schwenk et al., Nucleic Acids Res; 26(6):1427-32 (1998)). Furthermore, the generation of conditional mutants is a time consuming and labor intensive procedure, since the recombinase recognition strain and the recombinase expressing strain have to be breed at least over two generations in order to obtain animals carrying both, the recombinase transgene and two copies of the RRS-flanked target gene

[0004] Protein domains that have the ability to cross cell membranes were identified in the Antennapedia protein from Drosophila (Vives et al., J Biol Chem, 272(25):16010-7 (1997)), VP22 from HSV (Elliott and O'Hare, Cell, 88(2): 223-33 (1997)) and TAT from HIV (Green and Loewenstein, Cell, 55(6):1179-88 (1988); Frankel and Pabo, Cell, 55(6): 1189-93 (1988)). Fusion of such domains to heterologous proteins conferred the ability to transduce into cultured cells (Fawell et al., Proc Natl Acad Sci U S A, 91(2):664-8 (1994); Elliott and O'Hare (1997), Phelan et al., Nature Biotech. 16; 440-443 (1998) and Dilber et al., Gene Ther., 6(1):12-21 (1999)). Dalby and Bennett showed that a fusion protein consisting of VP22 and functional Fip recombinase translocated between cells in culture (from COS-1 cells transfected with VP22-Fip to CHO cells carrying Fip recognition sites (FRT sites); see Dalby and Bennett, Invitrogen, Expressions 6.2, page 13 (1999)).

[0005] On the other hand, a recent report demonstrated that the ß-galactosidase protein fused to the 11 amino acids transduction domain from the HIV TAT protein can infiltrate all tissues of living mice reaching every single cell (Schwarze et al., Science, 285(5433):1569-72 (1999)).

[0006] It was found that site-specific DNA recombinases can be translocated into cells of a living organism when fused to a protein transduction domain. Thus, whenever a gene mutation is desired, recombination is induced upon the injection of the appropriate site-specific recombinase fused to a transduction domain into such a living organism (provided, however, that said organism carries at least one appropriate RRS integrated in the genome).

[0007] The present invention thus provides

(1) the use of a fusion protein comprising

(a) a site-specific DNA recombinase domain and

(b) a protein transduction domain

for preparing an agent for inducing target gene alterations in a living organism, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in its genome;

- (2) a method for inducing gene alterations in a living organism which comprises administering to said living organism a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain as defined in (1) above, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in its genome;
  - (3) a fusion protein comprising

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- (a) a site-specific DNA recombinase domain and
- (b) a protein transduction domain

provided that when the site-specific DNA recombinase domain is wild type Flp then the protein transduction domain is not the VP22 protein of HSV (i.e., the fusion protein is not identical to the fusion protein of Dalby and Bennett (1999));

(4) a DNA sequence coding for the fusion protein of (3) above;

- (5) a vector comprising the DNA sequence as defined in (4) above;
- (6) a host cell transformed with the vector of (5) above and/or comprising the DNA of (4) above;
- (7) a method for producing the fusion protein of (1) above which comprises culturing the transformed host cell of
- (6) above and isolating the fusion protein; and
- (8) an injectable composition comprising the fusion protein as defined in (1) or (3) above.

[0008] The invention is further illustrated by the appended Figure and is explained in detail below.

[0009] Figure 1: Generation of induced mouse mutants using purified fusion proteins.

A: Expression of the fusion protein consisting of the site-specific DNA recombinase (e.g. Cre) and the protein transduction domain (e.g. the HIV derived TAT peptide) in prokaryontic or eukaryontic cells.

B: Extraction and purification of the expressed fusion protein (e.g. as described in Nagahara et al., 1998).

C: Injection of the purified fusion protein into mice carrying the RRS-flanked target sequence.

D: Analysis of the pattern of induced target gene recombination and the resulting phenotype. Triangle: RRS.

[0010] The expression "target sequences" according to the present invention means all kind of sequences which may be mutated (viz. deleted, translocated, integrated and/or inverted) by the action of the recombinase. The number of RRS in the target sequence depends on the kind of mutation to be performed by the recombinase. For most of the mutations (especially for deletions and invertions) two RRS are required which are flanking the sequence to be mutated (deleted or inverted). For some kinds of integrations only one RRS may be necessary within the target sequence.

[0011] The "living organisms" according to the present invention are multi-cell organisms and can be vertebrates such as mammals (e.g., rodents such as mice or rats) or non-mammals (e.g., fish) or can be invertebrates such as insects or worms. Most preferred living organisms are mice and fish.

[0012] The site-specific DNA recombinase domain within the fusion protein of the invention of the present application is preferably selected from a recombinase protein derived from Cre, Flp, ¢C31 recombinase (Thorpe and Smith, Proc. Natl. Acad. Sci, USA, vol. 95, 5505-5510 (1998)) and R recombinase (Araki et al., J. Mol. Biol., 182, 191-203 (1985)). The preferred recombinases are Cre (e.g., the Cre variant of aa 15 to 357 of SEQ ID NO: 2 or aa 325-667 of SEQ ID NO: 6) and Flpe (i.e., the Flp variant of aa 15 to 437 of SEQ ID NO: 4 or aa 325 to 747 of SEQ ID NO: 8).

[0013] The protein transduction domain according to the present invention is preferably derived from the Antennapedia protein of Drosophila, from the VP22 protein of HSV or from the TAT protein of HIV. Preferrably the protein transduction domain is derived from the TAT protein among which a TAT protein comprising the amino acid sequence shown in SEQ ID NO: 10 is most preferred.

[0014] The fusion of the two domains of the fusion protein can occur at any possible position, i.e., the protein transduction domain can be fused to the N- or C-terminal of the site-specific DNA recombinase or can be fused to active sites within the site-specific DNA recombinase. Preferrably the protein transfusion domain is fused to the N-terminal of the site-specific DNA recombinase domain.

[0015] The protein transduction domain can be fused to the site-specific DNA recombinase either through a direct chemical bond or through a linker molecule. Such linker molecule can be any bivalent chemical structure capable of linking the two domains. The preferred linker molecule according to the present invention is a short peptide, .g., having 1 to 20, preferrably 1 to 10, amino acid residues. Specifically preferred short peptides are ssentially consisting of Gly, Ala and/or Leu.

[0016] The fusion protein of the invention of the present application may further comprise other functional sequences

such as secretion conferring signals, nuclear localization signals and/or signals conferring protein stabilization.

[0017] In case the fusion protein comprises a protein transduction domain derived from the TAT protein of HIV, the DNA sequence coding for said fusion protein preferrably comprises the sequence

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[0018] Such a preferred DNA sequence is for instance shown in SEQ ID NO: 11. In said sequence the 3' terminal codon ggc codes for the linker Gly. The DNA sequence of a suitable recombinase may be directly attached to said codon ggc.

[0019] The fusion protein can be obtained by the following steps:

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- 1. Fusion of the recombinase coding region (e.g. encoding Cre: see amino acids 15 to 357 of SEQ ID NO: 2) with the sequence conferring protein translocation (e.g. the sequence encoding the TAT peptide YGRKKRRQRRR, SEQ ID NO: 10) using standard cloning protocols (Maniatis et al., Cold Spring Harbor Laboratory, New York (1989)) or chemical synthesis.
- 2. Generation of a construct for the expression of the fusion protein in prokaryotic or eukaryotic cells, e.g. in E. coli DH5a (Hanahan, J. Mol. Biol.;166(4):557-80 (1983)) using the QIAexpress pQE vector (Qiagen, Hilden).
- 3. Expression of the above mentioned fusion protein in prokaryotic or eukaryotic cells, e.g. in E. coli DH5a (Hanahan, 1983)
- 4. Extraction and purification of the above mentioned fusion protein e.g. as described in Nagahara et al., Nat. Med., 4(12):1449-52 (1998).
- [0020] Injection of the purified fusion protein into a living organism (e.g., a mouse) carrying a gene comprising the RRS-flanked target sequence (e.g., in an amount of 5 to 50 μg per g body weight). To demonstrate the feasibility of the invention, a reporter mouse strain is used harbouring a RRS-flanked flanked cassette which, when deleted by the recombinase, allows the expression of a cellular marker protein, such as β-galacosidase (Thorey et al., Mol. Cell Biol., 18(10):6164 (1998)).
- [0021] Analysis is achieved by determining the pattern of induced target gene recombination (e.g. through Southern blot analysis and X-Gal staining on tissue sections; Maniatis et al., 1989; Gossler and Zachgo, Joyner AL (Ed.), Oxford University Press, Oxford, New York (1993)).
  - [0022] The procedure's advantages over current technology are as follows:
- 35 (i) The absence of background recombination before administration of the fusion protein.
  - (ii) The reduction of time and resources which are necessary to combine the recombinase transgene and two copies of the RRS-flanked target gene by conventional breeding.

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#### Claims

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- Use of a fusion protein comprising
  - (a) a site-specific DNA recombinase domain and
  - (b) a protein transduction domain
  - for preparing an agent for inducing target gene alterations in a living organism, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in its genome.
  - 2. The use of claim 1 wherein the site-specific DNA recombinase domain is selected from a recombinase protein derived from Cre, Flp, φC31 recombinase, and R recombinase and preferably is Cre having amino acids 15 to 357 of SEQ ID NO: 2 or Flpe having amino acids 15 to 437 of SEQ ID NO: 4.
  - 3. The use of claim 1 or 2 wherein the protein transduction domain is a protein derived from the Antennapedia protein of Drosophila, from the VP22 protein of HSV or from the TAT protein of HIV, and preferably is derived from the TAT protein.
- 55 4. The use of claim 3, wherein the TAT protein comprises the amino acid sequenc

#### YGRKKRQRRR (SEQ ID NO: 10).

- The use of claims 1 to 4, wherein the protein transduction domain is fused to the N-terminal of the site-specific DNA recombinase domain.
  - 6. The use of claims 1 to 5, wherein the protein transduction domain is fused to the site-specific DNA recombinase domain through a direct chemical bond or through a linker molecule.
  - 7. The use of claim 6, wherein the linker molecule is a short peptide having 1 to 20, preferably 1 to 10 amino acid residues.
  - 8. The use of claims 1 to 7, wherein said fusion protein further comprises additional functional sequences.
  - 9. The use of claim 1, wherein the fusion protein has the sequence shown in SEQ ID NOs: 2, 4, 6 or 8.
  - 10. The use of claims 1 to 8, wherein the living organism is a vertebrate, preferably a rodent or a fish.
- 20 11. A method for inducing gene alterations in a living organism which comprises administering to said living organism, a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain as defined in claims 1 to 9, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in its genome.
- 25 12. A fusion protein comprising

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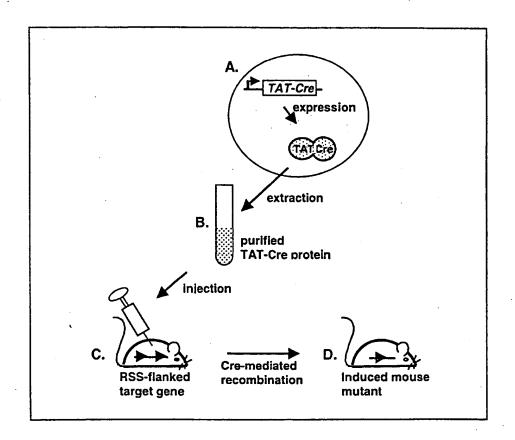
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- (a) a site-specific DNA recombinase domain and
- (b) a protein transduction domain provided that when (a) is Flp then (b) is not the VP22 protein of HSV.
- 13. The fusion protein of claim 11 being as defined in claims 2 to 9.
- 14. The fusion of claim 12 or 13, wherein the protein transduction domain is derived from the TAT protein of HIV.
- 35 15. A DNA sequence coding for the fusion protein of claim 12.
  - 16. The DNA sequence of claim 15 comprising the sequence shown in SEQ ID NOs: 9 and/or 11.
  - 17. A vector comprising the DNA sequence of claim 15.
  - 18. A host cell transformed with the vector of claim 17 and/or comprising the DNA of claim 15.
  - 19. A method for producing the fusion protein of claim 11 which comprises culturing the transformed host cell of claim 17 and isolating the fusion protein.
  - 20. An injectable composition comprising the fusion protein as defined in claims 1 to 9 or12 to14.

Fig. 1





#### PARTIAL EUROPEAN SEARCH REPORT

which under Rule 45 of the European Patent ConventionEP 00 10 0351 shall be considered, for the purposes of subsequent proceedings, as the European search report

				1					
	DOCUMENTS CONSID	ERED TO BE RELEVANT	r						
Category	Citation of document with in of relevant pass	ndication, where appropriate, ages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.7)					
X	WO 99 11809 A (IMP ;CRISANTI ANDREA (G 11 March 1999 (1999 * example 3 *		10-13,	C12N15/62 C12N9/00 C12N5/10 C12N1/21 C07K14/435					
X	WO 99 60142 A (HEND ;MARPLES BRIAN (GB) CANCE) 25 November * claim 9 *	; SCOTT SIMON (GB);	1-3, 5-13,15, 17-20	C07K14/035					
D,X	Translocation" EXPRESSIONS,	er(TM) - The power of	13						
	vol. 6, no. 1, Febr page 6 XPO02140132 * column 1, paragra	uary 1999 (1999-02), ph 7 *							
A	SCHWARZE S ET AL: transduction: deliv active protein into	ery of a biologically							
i	SCIENCE., vol. 285, no. 5433,			TECHNICAL FIELDS SEARCHED (Int.Cl.7)					
·	3 September 1999 (1 1569-1572, XP002140	999-09-03), pages		C12N C07K A01K A61K					
INCO	MPLETE SEARCH	······································	<u> </u>	·					
not comp be carried Claims se Claims se	ch Division considers that the present by with the EPC to such an extent that out, or can only be carried out partia sarched completely: sarched incompletely;	application, or one or more of its claims, does a meaningful search into the state of the art of ity, for these claims.	s/do Jannol						
Alt	the human/animal bod	rected to a method of to y (Article 52(4) EPC),	the						
sea eff	rch has been carried ects of the compound	out and based on the a composition.	i regea						
	Place of search	Date of completion of the search	<u> </u>	Examiner					
	THE HAGUE	14 June 2000	Lon	noy, 0					
X : par Y : par doo A : tec	ATEGORY OF CITED DOCUMENTS ticularly relevant if taken alone ticularly relevant if combined with ano ument of the same category thrological background	E : earlier patent do after the filing da ther D : document ched L : document ched	cument, but publite in the application or other reasons	lahed on, or					
O ; no	n-written disclosure ermediate document	& ; member of the s document	& : member of the same patent family, corresponding						

EPO FORM 1509 03.82 (P04C07)



Application Number

EP 00 10 0351

CLAIMS INCURRING FEES
The present European patent application comprised at the time of filling more than ten claims.
Only part of the claims have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims and for those claims for which claims fees have been paid, namely claim(s):
No claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims.
LACK OF UNITY OF INVENTION
The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:
·
see sheet B
All further search fees have been paid within the fixed time limit. The present European search report has been drawn up for all claims.
As all searchable claims could be searched without effort justifying an additional fee, the Search Division did not invite payment of any additional fee.
Only part of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the inventions in respect of which search fees have been paid, namely claims:
None of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the invention first mentioned in the claims, namely claims:



#### PARTIAL EUROPEAN SEARCH REPORT

Application Number

EP 00 10 0351

	OOCUMENTS CONSIDERED TO BE RELEVANT	CLASSIFICATION OF THE APPLICATION (Int.CL7)	
ategory	Citation of document with Indication, where appropriate, of relevant passages	Relevant to claim	
A	WO 95 00555 A (EUROP MOLECULAR BIOLOGY LAB EM ;STEWART FRANCIS (DE)) 5 January 1995 (1995-01-05)		
	·		
			TECHNICAL FIELDS SEARCHED (Int.CL7)
		·	



# LACK OF UNITY OF INVENTION SHEET B

Application Number EP 00 10 0351

The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

1. Claims: 1-3,5-8,10-13,15,17-20 (all partially)

Use of a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain for preparing an agent for inducing target gene alterations in a living organism carrying at least one recognition site in its genome; said use wherein the site-specific recombinase is the Cre recombinase and said protein transduction domain is derived from Antennapedia protein of Drosophila; corresponding method, fusion protein, DNA sequence, vector, host cell and composition.

2. Claims: 1-3,5-13,15,17-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the Cre recombinase and said protein transduction domain is derived from VP22 of HSV, eventually as presented in SeqIdNo.6

3. Claims: 1-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the Cre recombinase and said protein transduction domain is derived from Tat of HIV, eventually as presented in SeqIdNo.2, eventually comprising the Tat-derived sequences of SeqIdNo.9, SeqIdNo.10 or SeqIdNo.11

4. Claims: 1-3,5-8,10-13,15,17-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the Flp recombinase or its modified variant Flpe, and said protein transduction domain is derived from AntP of Drosophila

5. Claims: 1-3,5-13,15,17-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the Flp recombinase or its modified variant Flpe, and said protein transduction domain is derived from VP22 of HSV, eventually as presented in SeqIdNo.8

6. Claims: 1-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the Flp recombinase or its modified variant Flpe, and said protein transduction domain is derived from Tat of HIV, eventually as presented in SeqIdNo.4, eventually comprising



#### LACK OF UNITY OF INVENTION SHEET B

Application Number

EP 00-10 0351

The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

the Tat-derived sequences of SeqIdNo.9, SeqIdNo.10 or SeqIdNo.11

7. Claims: 1-3,5-8,10-13,15,17-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the PhiC31 recombinase, and said protein transduction domain is derived from AntP of Drosophila

8. Claims: 1-3,5-8,10-13,15,17-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the PhiC31 recombinase, and said protein transduction domain is derived from VP22 of HSV

9. Claims: 1-8,10-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the PhiC31 recombinase, and said protein transduction domain is derived from Tat of HIV, eventually comprising the Tat-derived sequences of SeqIdNo.9, SeqIdNo.10 or SeqIdNo.11

10. Claims: 1-3,5-8,10-13,15,17-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the R recombinase, and said protein transduction domain is derived from AntP of Drosophila

- 11. Claims: 1-3,5-8,10-13,15,17-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the R recombinase, and said protein transduction domain is derived from VP22 of HSV

12. Claims: 1-8,10-20 (all partially)

As for subject 1, but wherein said site-specific recombinase is the R recombinase, and said protein transduction domain is derived from Tat of HIV, eventually comprising the Tat-derived sequences of SeqIdNo.9, SeqIdNo.10 or SeqIdNo.11

#### ANNEX TO THE EUROPEAN SEARCH REPORT ON EUROPEAN PATENT APPLICATION NO.

EP 00 10 0351

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report. The members are as contained in the European Patent Office EDP file on The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

14-06-2000

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WO	9500555	A	05-01-1995	EP AT AU CA DE DE EP ES JP US	0632054 A 152123 T 7227994 A 2166198 A 69402863 D 69402863 T 0707599 A 2101552 T 2986915 B 8511681 T 6040430 A	04-01-1999 15-05-199 17-01-1999 05-01-1999 28-05-1999 31-07-1999 24-04-1999 01-07-1999 10-12-1999 21-03-2000
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For more details about this annex : see Official Journal of the European Patent Office, No. 12/82